Tetrahedron Letters, Vol.27, No.2, pp 143-146, 1986 0040-4039/86 \$3.00 + .00 Printed in Great Britain ©1986 Pergamon Press Ltd.

> Application of Long Range 1 H/ 13 C Heteronuclear Correlation Spectroscopy (LR HETCOSY) to Structure Elucidation: The Structure of Murayaquinone

Yukiharu Sato[†], Rodger Kohnert, Steven J. Gould^{*,1} Department of Chemistry, Oregon State University Corvallis, Oregon 97331

> [†]On leave from Tamagawa University, 6-1-1 Tamagawa Gakuen, Machida-shi, Tokyo 194, Japan

<u>Abstract:</u> The structure of a polyketide-derived, <u>o</u>-phenanthraquinone streptomyces metabolite has been deduced from interpretation of a 2D heteronuclear correlation experiment that reveals long range 1 H- 13 C couplings.

<u>Streptomyces murayamaensis</u> has previously been shown to produce the structurally unusual kinamycins A - D,² possessing a benzo[b]tetrahydrocarbazole skeleton.³ In the course of our studies of kinamycin biosynthesis,⁴ we observed two additional colored metabolites upon TLC analysis (silica gel) of crude extracts of <u>S</u>. <u>murayamaensis</u> fermentations. One of these, giving a pink spot on TLC, has now been characterized, primarily by the application of a simple 2D NMR experiment (LR HETCOSY) that reveals long-range ¹H-¹³C spin couplings.^{5,6} This experiment allowed the assignment of all the quaternary ¹³C resonances and defined the carbon-carbon connectivity for the structure of murayaquinone, 1, possessing the rare <u>ortho</u>-phenanthraquinone skeleton.

<u>S. murayamaensis</u> was grown as previously described, and the broth and mycelial mat were extracted with benzene.⁴ The extracts were dried (Na_2SO_4) and concentrated in vacuo to yield a gum (900 mg from 1.2 L) that was taken up in CHCl₃ and chromatographed on a column (2.2 x 33 cm) of Silicar CC-4 (Mallinkrodt 7086) prepared and eluted with CHCl₃. The pink band was collected separately (104 mg) and further purified by HPLC (two Partisil PXS 10/25 columns in series, iso-octane:THF = 9:1, 3.0 mL/min at 1600 psi, detection by refractive index). After removing the solvent, crystallization from CH₂Cl₂ yielded 9 mg of 1 : mp 108-109^oC.⁷

The infrared spectrum contained absorbances at 3400 (H-bonded OH), 1700 (ary]





ketone) and 1615 (quinone) cm⁻¹. A single quinone absorbance indicated a similar environment for each of these carbonyls. The ¹H NMR and ¹H COSY spectra (400.13 MHz) indicated one methyl group and one <u>n</u>-propyl group on quaternary unsaturated carbons, two hydrogen-bonded phenols, an isolated aromatic proton (penta-substituted ring) and a grouping of three aromatic protons indicative of 1,2,3-substitution. Analysis of the ¹³C NMR and ¹³C DEPT spectra (100.6 MHz) confirmed the two CH₃'s, two CH₂'s, and four CH's, as well as revealed eleven unsaturated quaternary carbons. Thus, the data suggested a tricyclic quinone with methyl and butyryl side chains. These substituents were inferred to be <u>ortho</u> to each other on the assumption that the biogenesis of <u>1</u> was from a straight-chain polyketide precursor. With the required hydrogen-bonding of phenols to both quinone carbonyls, four anthraquinones and four phenanthraquinones remained as possible structures.

All but one of these eight structures could be eliminated by data from the LR HETCOSY spectrum (Figure 1). The cross-peak matrices (2 x 4, 2 x 3, and 2 x 2) identify sets of directly bonded carbons and hydrogens. The remaining cross peaks were due to long range ${}^{1}\text{H}$ - ${}^{13}\text{C}$ coupling. Thus, the phenolic proton at 12.27 ppm is coupled to a methine carbon and two quaternary carbons, while the phenolic proton at 12.73 ppm shows coupling to three quaternary carbons. These data implied that the lone aromatic hydrogen (7.31 ppm) must be <u>para</u> to the latter phenol and <u>meta</u> to the methyl group (coupling between 2.20 and 126.2 ppm).

Four cross peaks with the aromatic hydrogen singlet at 7.31 ppm were observed. A cross peak from 126.2 ppm $({}^{3}J_{CH})$ demonstrated its <u>meta</u> relationship to the methyl group, thus confirming its <u>para</u> relationship to the 12.73 ppm phenol. A second cross peak, from 205.4 ppm, placed the butyryl side chain on the adjacent ring carbon, confirming the biogenetic assumption (vide infra), and a cross peak from 114.8 ppm confirmed this as the ring fusion next to the quinone. The most important cross peak was that from 135.0 ppm. Since this is not due to a quinone carbonyl, 1 must have a phenanthrene skeleton, and the structure shown fulfills all the requirements. All remaining cross peaks fully support the unique structure 1.

Only three polyketide-derived phenanthrenes $(2^8, 3^9, and 4^{10})$ have been previously characterized. The present example demonstrates how effective the LR HETCOSY can be in assigning quaternary carbons and mapping carbon-carbon connectivities.



Figure 1. Contour plot of 1 H/ 13 C LR HETCOSY spectrum of 1. Spectral aquisition parameters: 21739 Hz sweep width in the F₂ dimension; 256 spectra (128 scans each) were accumulated with a 1.0 sec relaxation delay and 0.179 msec increments across the interval 8.0 to 53.722 msec. Resolution was 21.2 Hz/pt in the F₂ dimension and 10.9 Hz/pt in the F₁ dimension.

<u>Acknowledgments</u>: Professor S. Ōmura is thanked for providing a culture of <u>S</u>. <u>muray-amaensis</u>. This work was supported by Public Health Research Grant GM31715 to S.J.G. NMR Spectra were obtained on a Bruker AM 400 spectrometer purchased in part through grants from the National Science Foundation (PCM-8216190) and from the M. J. Murdock Charitable Trust, and the high resolution mass spectrum was obtained on a Kratos MS 50 TC spectrometer purchased with grants from the National Institutes of Health Division of Research Resources (DRR 1S10RR01409) and from the Anheuser-Busch Company.

<u>References:</u>

- 1. Career Cevelopment Awardee of the National Cancer Institute (CA 00880), 1979-1984.
- S. Ito, T. Matsuya, S. Omura, M. Otani, A. Nakagawa, H. Takeshima, Y. Iwai, and T. Hata, <u>J. Antibiot.</u>, 23, 315 (1970); T Hata, S. Omura, Y. Iwai, A. Nakagawa, M. Otani, S. Ito, and T. Matsuya, <u>ibid.</u>, 24, 353 (1971).
- S. Ōmura, A. Nakagawa, H. Yamada, T. Hata, A. Furusaki, and T. Watanabe, <u>Chem.</u> <u>Pharm. Bull.</u>, <u>19</u>, 2428 (1971); S. Ōmura, A. Nakagawa, H. Yamada, T. Hata, A. Furusaki, and T. Watanabe, <u>ibid.</u>, 21, 931 (1973).
- 4. Y. Sato and S. J. Gould, <u>Tetrahedron Lett</u>. <u>26</u>, 4023 (1985).
- The pulse pulse sequence provides both one-bond and long range heteronuclear couplings, and is described in H. Bleich, S. Gould, P.Pitner, and J. Wilde, <u>J. Mag.</u> <u>Reson.</u>, <u>56</u>, 515 (1984). For an example of its use for ¹H/¹³C couplings see Y. 7Sato, M. Geckle, and S. J. Gould, <u>Tetrahedron Lett.</u>, 26, 4019 (1985).
- Recently, other approaches for detecting long range J_{CH}'s have been published: H. Seto, K.Furihata, N. Otake, Y. Itoh, S. Takahashi, Tl Haneishi, and M. Ohuchi, <u>Tetrahedron Lett.</u>, 25, 337 (1984); H. Kessler, C. Griesinger, and J. Lautz, <u>Angew. Chem. Int. Ed. Engl.</u>, 23, 444 (1984); W. F. Reynolds, R. G. Enriquez, L. I., and Escobar, and X. Lozoya, <u>Can. J. Chem.</u>, 62, 2421 (1984); and J. P. Kintzinger, P. Maltese M.
 - Bourdonneau, and C. Brevard, Tetrahedron Lett., 25, 6007 (1984).
- 7. UVmax (MeOH) 524 (ε =4485), 401 (ε =3175), 288 (ε =13920), 242 nm (ε =18284); ¹H NMR (CDC1₃, 400.13 MHz) [§]1.00 (3H, t, J = 7.5 Hz), 1.71 (2H, m), 2.20 (3H, s), 2.78 (2H, t, J = 7.5 Hz), 6.94 (1H, d, J = 8.6 Hz), 7.31 (1H, s), 7.40 (1H, d, J = 8.3 Hz), 7.54 (1H, dd, J = 8.6 and 8.3 Hz), 12.27 (1H, s), 12.72 (1H, s); ¹³C NMR (CDC1₃, 100.6 MHz) [§]205.4 (s), 182.6 (s), 182.2 (s), 166.1 (s), 165.0 (s), 150.0 s), 139.4 (d), 135.0 (s), 132.8 (s), 126.2 (s), 119.6 (d), 115.9 (d), 115.3 (s), 114.8 (s), 113.1 (d), 44.9 (t), 17.3 (t), 13.8 (q), 12.0 (q); HRMS (70 ev) m/z calcd. for C₁₉H₁₆O₅: 324.0097, fnd. 324.1006.
- J. Polonsky, B. C. Johnson, P. Cohen, and E. Lederer, <u>Bull. Soc. Chim. Fr.</u>, 1909 (1963).
- 9. M. Lounasmaa and J. Zylber, *ibid.*, 3100 (1969).
- 10. B. Krone, A. Hinrichs, and A. Zeeck, <u>J. Antibiot.</u>, 34, 1538 (1981).

(Received in USA 21 May 1985)